

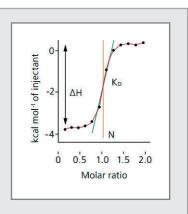
Calorimetric system – Auto-iTC 200

Isothermal Titration Calorimetry (ITC)

Calorimeter is designed to measure the heat of binding. In a typical arrangement, the titrant, also referred as the ligand, is injected into the sample cell containing the macromolecule sample solution. The calorimetric measurement can be done over a range of biologically relevant conditions (temperature, salt, pH, etc.). No labelling is necessary and the complete thermodynamic profile of the interaction can be obtained in a single measurement. ITC system directly measure submillimolar to nanomolar binding constants (10³ - 10⁵ M⁻¹). The interactions with nanomolar to picomolar binding constants (10⁵-10¹² M⁻¹) can be measured using the competitive binding technique, the same principle can be used for low affinity interactions (10³-10² M⁻¹).

ITC method can be used for

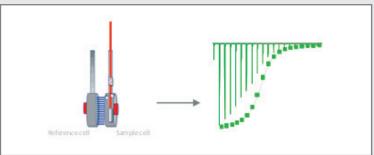
- characterization of biomolecular interactions of small molecules, proteins antibodies, nucleic acids, lipids and others
- enzyme kinetics studies, biological activity or the effect of molecular structure changes on binding mechanism determination
- determination of thermodynamic parameters K_A , ΔH and ΔS values, stoichiometry or kinetics parameters K_A and K_{CAT}



Technical Specifications

Instrument: Auto-iTC200 (Malvern)





Features:

- operating temperature range is of 2°C to 80°C
- sample tray storage temperature range: 4°C to ambient
- coin-shaped calorimetric cell (Hastelloy)

Operational mode:

calorimetric measurement is performed by core facility staff only

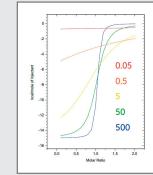
Data evaluation software: Origin software, NITPIC (possibility to train people in data processing)

Established Methodologies and Provided Services

- calorimetric measurement of protein-ligand interaction (Standard titration method, Single injection method) or competitive-based measurement low (10³ M⁻¹) or high (10⁹ M⁻¹) affinity interactions
- data evaluation thermodynamic parameters determination using curve fitting models: One set of binding sites,
 Two sets of binding sites
- eventuality of manual data evaluation using fitting models: Sequential binding sites, Competitive binding, Dissociation (data evaluation assistance)
- · basic ITC data evaluation training

Sample Requirements

- **Proper sample preparation is crucial** for the successful ITC measurement. The buffer solution, in which the macromolecule and the ligand are dissolved, **should be exactly the same** (dialysis or lyophilisation and dissolution in the buffer for ITC). The pH should be checked before the measurement.
- The macromolecule sample (the sample placed in the cell): 450 μl
- The ligand solution (the sample placed in the injection syringe): 150 µl
- Sample concentrations must be determined precisely.
- Generally a concentration of ligand should be 10 times higher than the concentration of macromolecule otherwise the concentration should be optimized.
- High affinity interactions can be studied at low concentrations. In this case the minimum concentration of
 macromolecule sample which causes measurable heat is 10 μM. For low affinity interactions the macromolecule
 sample concentration should be 5 times of K_D or higher, but higher concentration may be limited by availability or
 solubility of samples.
- Calculating the cell sample concentration M = c / (n x K_A) = c / n x K_D
 M ... molar concentration of the cell sample; c-value ... should lie between 10-500;
 n ... binding stoichiometry; K_A ... association constant; K_D ... dissociation constant
- At least 10 ml of the used buffer must be sent for each measurement (for AutoiTC200).
- If it is possible, choose a pH buffer with low heat ionization in order to minimize artifactual heats of buffer ionization.
- If the presence of reducing agent is required for a protein stability, then ß-mercaptoethanol (less than 5 mM) or TCEP (less than 2 mM) should be used rather than DTT.



The shape of the binding isotherms depend on a unitless constant c

It is recommended to discuss the project and the details of the experiment (sample preparation, sample requirements) with the Core Facility members in advance.

Contacts

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Instrument Location:

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